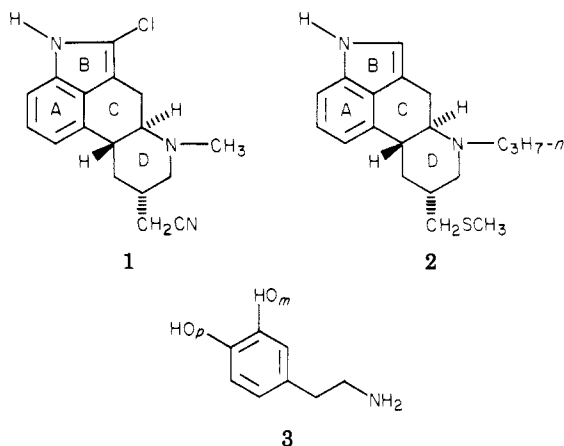


Communications to the Editor

Proposed Dopaminergic Pharmacophore of Lergotriple, Pergolide, and Related Ergot Alkaloid Derivatives

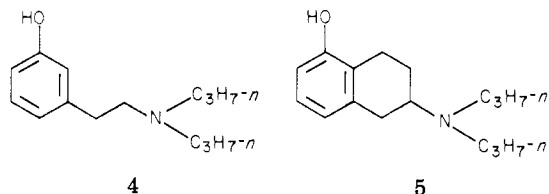
Sir:

The central dopamine agonist properties of certain derivatives of ergot alkaloids, typified by lergotriple (1) and

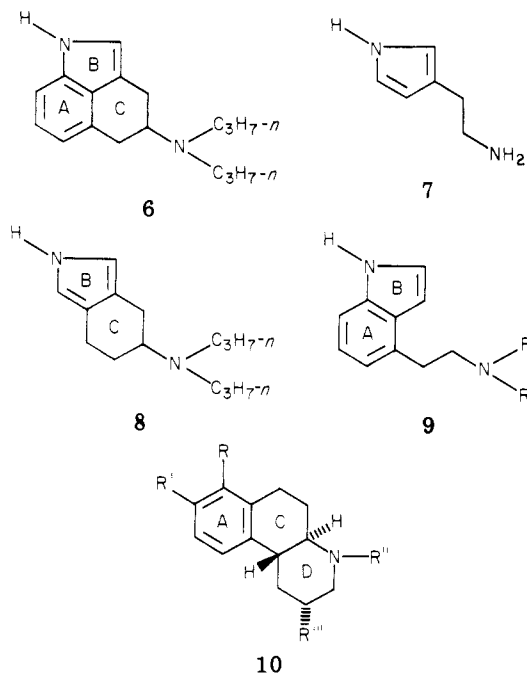


pergolide (2), have been well documented in the literature.^{1,2} Marek and Roth³ stated that 1 is a potent agonist at presynaptic dopamine receptors on striatal and mesolimbic nerve terminals. Goldstein et al.⁴ concluded that lergotriple has the properties in the CNS of a mixed agonist-antagonist with respect to some presynaptic dopamine receptors. However, there seems to be little structural resemblance between 1 and 2 and dopamine (3), and a reviewer⁵ stated in 1978 that ergot alkaloids bear little structural resemblance to the dopaminergic aporphines and 2-aminotetralins.

In 1978, one of us⁶ suggested that lergotriple is related structurally to dopamine, if it is accepted that the weakly acidic indole NH group is bioisosteric with the "meta" OH of dopamine. The meta OH has been proposed⁷ to be of considerable importance in agonist-receptor interaction. Some support is given to this proposal by the report of Geissler⁸ that *N,N*-di-*n*-propyl-*m*-tyramine (4) has dop-



amine agonist actions and the report of McDermed et al.⁹ that 2-(di-*n*-propylamino)-5-hydroxytetralin (5) is a potent dopaminergic. Bach and Kornfeld¹⁰ described the ergoline fragment 6 and stated that it inhibited prolactin secretion



and dopamine binding, which are characteristic actions of dopaminergic agonists. Bach et al.¹¹ found that 3-(2-aminoethyl)pyrrole (7) was ineffective in lowering prolactin levels, which was attributed to rapid metabolic inactivation of the primary amine by monoamine oxidase. No tertiary amine derivative of 7 was reported. Compound 8, a BC bicyclic ergoline partial structure, exhibited prolactin inhibitory activity, as well as some activity in a rat rotation assay, albeit in high doses in both assays. Bach et al.¹² have reported that depyrroloergolines (10, R = R' = H; R'' = *n*-C₃H₇; R''' = CH₂SCH₃) are dopaminergically inactive in two tests. In contrast, catechol derivatives of 10 (R = R' = OH; R'' = alkyl; R''' = H) are highly active, potent dopaminergics.⁶

Inspection of the molecular structures of lergotriple (1) and pergolide (2) suggested that the pharmacophore of lergotriple, pergolide, and 6 may be a 4-(2-aminoethyl)indole system, 9. Hofmann and Troxler¹³ described derivatives of 9 (R, R' = H, Me) and suggested that these "could be used in treatment of asthma". However, the literature has not revealed that they have been evaluated for dop-

- (1) McDermed, J.; Miller, R. J. *Annu. Rep. Med. Chem.* **1979**, *14*, 12.
- (2) Fuller, R. W.; Clemens, J. A.; Kornfeld, E. C.; Snoddy, H. R.; Smalstig, E. B.; Bach, N. J. *Life Sci.* **1979**, *24*, 375.
- (3) Marek, K. L.; Roth, R. H. *Eur. J. Pharmacol.* **1980**, *62*, 137.
- (4) Goldstein, M.; Lew, J. Y.; Nakamura, S.; Battista, A. F.; Lieberman, A.; Fuxe, K. *Fed. Proc., Fed. Am. Soc. Exp. Biol.* **1979**, *37*, 2202.
- (5) McDermed, J.; Miller, R. J. *Annu. Rep. Med. Chem.* **1978**, *13*, 11.
- (6) Cannon, J. G. In "Peripheral Dopaminergic Receptors"; Imbs, J.-L.; Schwartz, J., Eds.; Pergamon Press: New York, 1979; pp 87-94.
- (7) Cannon, J. G. *Adv. Neurol.* **1975**, *9*, 177.

- (8) Geissler, H. E. *Arch. Pharm. (Weinheim)* **1977**, *310*, 749.
- (9) McDermed, J. D.; McKenzie, G. M.; Freeman, H. S. *J. Med. Chem.* **1976**, *19*, 547.
- (10) Bach, N. J.; Kornfeld, E. C. U.S. Patent 4110339, 1978; *Chem. Abstr.* **1979**, *90*, 121415f.
- (11) Bach, N. J.; Kornfeld, E. C.; Jones, N. D.; Chaney, M. O.; Dorman, D. E.; Paschal, J. W.; Clemens, J. A.; Smalstig, E. B. *J. Med. Chem.* **1980**, *23*, 481.
- (12) Bach, N. J.; Kornfeld, E. C.; Clemens, J. A.; Smalstig, E. B. *J. Med. Chem.* **1980**, *23*, 812.
- (13) Hofmann, A.; Troxler, F. French Patent 1344579, 1963; *Chem. Abstr.* **1964**, *60*, 11988.

Table I. Sympathetic Neuronal Inhibiting Activity of Lergotriole (1), Pergolide (2), and 4-[2-(Di-*n*-Propylamino)ethyl]indole (9) in Anesthetized Cats

no.	dose, $\mu\text{g}/\text{kg}$	% inhibn of cardioaccelerator nerve stimulation, 2 Hz	inhibitory effect reversed by haloperidol, 100 $\mu\text{g}/\text{kg}$	ID ₅₀ , $\mu\text{mol}/\text{kg}$ (95% CL)	potency ratio ^a rel to apomorphine (fiducial limits)
1	30	21 \pm 7	yes, $p < 0.01$, $n = 5$	0.27 (0.13-0.62)	0.08 (0.04-0.16)
	100	57 \pm 12			
	300	69 \pm 10			
2	3	23 \pm 3	yes, $p < 0.01$, $n = 7$	0.02 (0.01-0.02)	1.13 (0.7-1.8)
	10	59 \pm 6			
	30	80 \pm 5			
9	30	29 \pm 10	yes, $p < 0.01$, $n = 7$	0.22 (0.12-0.33)	0.14 (0.07-0.33)
	100	60 \pm 4			
	300	77 \pm 8			

^a Apomorphine was used as a reference dopamine agonist inhibiting the positive chronotropic response induced by stimulation of the right cardioaccelerator nerve at 2-Hz frequency.

amine-like effects. In the present work, the target compound based on 9 bears *n*-propyl groups on the side-chain amino group, consistent with several reports⁶ that this N-substitution tends to maximize dopaminergic activity and potency.

Synthesis of 9 ($R = R' = n\text{-C}_3\text{H}_7$) began with a Reissert indole synthesis using 6-chloro-2-nitrotoluene. The 4-chloroindole product was converted into 4-cyanoindole with $\text{Cu}_2(\text{CN})_2$.¹⁴ This nitrile was hydrolyzed to indole-4-carboxylic acid, and the *N*-benzoyl derivative of this acid was homologated to indole-4-acetic acid by an Arndt-Eistert sequence. This carboxylic acid was converted to its *N,N*-di-*n*-propylamide with di-*n*-propylamine and hexamethylphosphorous triamide/ CCl_4 . The amide was reduced with LiAlH_4 to 9 ($R = R' = n\text{-C}_3\text{H}_7$), which was characterized and evaluated biologically as its bifumarate salt, mp 154-155 °C (EtOH-Et₂O-petroleum ether): MS, m/e 244 (M^+ - fumaric acid).

Pharmacology. Methods. Inhibition of Postganglionic Cardioaccelerator Nerve in Cats. Anesthesia was induced by intrathorax administration of pentobarbital sodium (30 mg/kg). Arterial pressure was measured from the right femoral artery using a Statham P23AA pressure transducer and was recorded using a Beckman RS dynograph. The pulses were integrated and recorded by use of a cardiometer. The respiration was supported by a Harvard respiratory pump and, following a midline incision of the thorax, bipolar platinum electrodes were placed on the right postganglionic cardioaccelerator nerves for stimulation using a Grass S4S stimulator. The frequency of stimulation was 2 Hz. The impulses were delivered from 20-30 s and a pulse duration of 5 ms was used. Supramaximal voltage was used. After the establishment of consistent controls, 1, 2, or 9 was administered to cats in doses that varied by 0.48 log intervals. The ability of 1, 2, and 9 to affect mean arterial pressure and resting heart rate was determined, as well as the ability to inhibit neuronal sympathetic transmission. At the completion of each experiment, 100 μg of haloperidol was administered iv, and the ability of the compounds to inhibit cardioaccelerator nerve stimulation was redetermined.

Inhibition of Spontaneous Locomotion in Rats. Sprague-Dawley rats (225-250 g) were kept in a lighted room for at least 24 h. Pairs of rats were injected sc with 9 ($R = R' = n\text{-C}_3\text{H}_7$) and placed in a darkened room for 30 min and then in a Plexiglas container, and locomotion was followed using an electromagnetic activity meter (Columbus Instruments, Model S) for an additional 30

min. The counts recorded during the first 6 min were discarded, and the following 24-min counts were recorded. Six pairs of rats were used for each dose and received either saline (0.9%) or 0.33, 1.0, or 3.3 mg/kg 9 in normal saline.

Renal Vasodilatation Experiments in Dogs. The standard procedure of Goldberg¹⁵ was used for studying the vascular (postsynaptic) effects of the compounds. Pentobarbital-anesthetized dogs were prepared for recording carotid blood pressure and renal artery flow. Various doses of the compounds and dopamine were injected intraarterially in a fixed volume of 0.2 mL.

Statistics. The ID₅₀ values for inhibition of cardioaccelerator nerve stimulation and inhibition of locomotion were calculated by the method for probit analysis, and bioassays were analyzed by parallel line assay as described by Finney.¹⁶

Results and Discussion

The presence of presynaptic dopaminergic receptors inhibiting sympathetic transmission has been well documented.¹⁷ Many compounds possessing dopamine-like properties have been shown to inhibit the positive chronotropic response induced by low-frequency nerve stimulation.¹⁸⁻²³ This neuronal activity involves the stimulation of presynaptic dopaminergic receptors present in the peripheral sympathetic nerve endings, resulting in a decrease of norepinephrine release, an effect blocked by dopamine antagonists. This has been used as a tool for detecting compounds exerting dopaminergic activity.

All of the compounds studied elicited significant dopaminergic activity. Table I shows that lergotriole (1) and 9 were equally active in anesthetized cats in a test for their ability to inhibit neuronal postganglionic cardioaccelerator nerve stimulation. Pergolide (2) was the most active in

(14) Uhle, F. C. *J. Am. Chem. Soc.* 1949, 71, 761.

- (15) Goldberg, L. I.; Volkman, P. H.; Kohli, J. D. *Annu. Rev. Pharmacol. Toxicol.* 1978, 18, 57.
 (16) Finney, D. J. "Statistical Methods in Biological Assays"; Griffin: London, 1952; pp 139-145.
 (17) Langer, S. Z. In "Presynaptic Receptors"; Langer, S. Z.; Starke, K.; Dubocovich, M. L., Eds.; Pergamon Press: New York, 1979; pp 13-22.
 (18) Ilhan, M.; Long, J. P.; Cannon, J. G. *Eur. J. Pharmacol.* 1975, 33, 13.
 (19) Ilhan, M.; Long, J. P. *Arch. Int. Pharmacodyn. Ther.* 1975, 216, 4.
 (20) Ilhan, M.; Long, J. P.; Cannon, J. G. *Arch. Int. Pharmacodyn. Ther.* 1976, 222, 70.
 (21) Sharabi, F. M.; Long, J. P.; Cannon, J. G.; Hatheway, G. J. *J. Pharmacol. Exp. Ther.* 1976, 199, 630.
 (22) Cannon, J. G.; Hatheway, G. J.; Long, J. P.; Sharabi, F. M. *J. Med. Chem.* 1976, 19, 987.
 (23) Sharabi, F. M.; Long, J. P.; Cannon, J. G. *J. Pharmacol. Exp. Ther.* 1977, 202, 97.

Table II. Cardiovascular Effects of Lergotriole (1), Pergolide (2), and 4-[2-Di-*n*-(Propylamino)ethyl]indole (9) in the Anesthetized Cat

no.	dose, $\mu\text{g}/\text{kg}$	effect on mean arterial blood pressure		
		% increase from resting control ^a	% decrease from resting control	% decrease in heart rate
1	30		22 \pm 4	11 \pm 3
	100		34 \pm 5	10 \pm 4
			(97 \pm 7) ^b	(152 \pm 8)
2	3	5 \pm 2	5 \pm 2	23 \pm 3
	10	15 \pm 4	8 \pm 2	59 \pm 6
9	30	52 \pm 17	6 \pm 2	80 \pm 5
	30	6 \pm 2	11 \pm 4	7 \pm 2
	100	16 \pm 2	18 \pm 5	12 \pm 2
			(101 \pm 4)	(123 \pm 13)
	300	41 \pm 21	10 \pm 4	10 \pm 4

^a The primary pressor effect induced by pergolide (2) and 9 was transient and of short duration. ^b Values within parentheses indicate resting mean arterial blood pressure (mmHg) and resting heart rate (beats/min).

this respect. All compounds lowered mean arterial pressure and resting heart rate. Compound 9 did not antagonize the positive chronotropic responses to epinephrine or isoproterenol, nor did 9 antagonize the vasopressor response to epinephrine. Haloperidol (100 $\mu\text{g}/\text{kg}$) antagonized significantly the inhibition of the cardioaccelerator nerve preparation induced by 1, 2, and 9. In most preparations, the heart-rate reductions and inhibition of neural stimulation induced by 1 and 2 lasted 60 min. Comparable doses of 9 lasted 2–3 h or more.

In rats, 9 induced decreased locomotion during the 24-min period that was measured 6 min after sc administration. The ED₅₀ ($\mu\text{mol}/\text{kg}$) and 95% confidence limits are 2.0 (0.33–4.1).

Following iv injection into cats, all compounds studied produced lowering of mean blood pressure, decrease in heart rate (Table II), and inhibition of positive chronotropic responses induced by stimulation of postganglionic fibers of the cardioaccelerator nerves. Compounds 1 and 9 required 30–40 min following iv administration to reach maximal inhibition of neural transmission. This slow rate of onset of action may indicate metabolic activation. The duration of effect of 9 was considerably greater than for

1 and 2. The inhibition by 9 of sympathetic neural transmission to the hearts, as well as the production of bradycardia, was not related to inhibition of β_1 receptors in the heart. The positive chronotropic responses induced by epinephrine or isoproterenol were not inhibited by 9. Likewise, 9 did not antagonize the vasopressor responses to epinephrine, indicating no inhibition of α_1 adrenoceptors of the arterial bed. Haloperidol (100 $\mu\text{g}/\text{kg}$) reversed neural inhibition produced by all compounds, which is evidence that they interact with dopaminergic inhibitory receptors on the adrenergic nerve terminal of cats.

Compound 9 induced decreased locomotion in rats. Whether this reflects synaptic dopaminergic neuronal inhibition in the striatum, or some other mechanism, has not been determined. However, this demonstrates the activity of 9 in a second animal species and indicates that it apparently crosses the blood–brain barrier.

None of the compounds produced renal vasodilatation, in accord with the report of McNay et al.²⁴ that lergotriole (1) does not dilate the renal vascular bed.

The biological data presented in this communication suggest that 9 ($R = R' = n\text{-C}_3\text{H}_7$) is a dopaminergic agonist. In the anesthetized cat, lergotriole (1) and 9 are quite parallel in their actions and potencies. The data are consistent with the proposal that the structure of 9 is the active pharmacophore in the lergotriole and pergolide molecules. Bach et al.¹¹ have proposed that the dopaminergic pharmacophore of ergoline is the pyrroleethylamine moiety. This contrast remains to be clarified by further studies.

Acknowledgment. This work was supported by Grant GM-22365 from the National Institute of General Medical Sciences. Thanks are extended to Dr. E. C. Kornfeld and Eli Lilly & Co. for a generous gift of pergolide.

(24) McNay, J. L.; McDonald, R. H., Jr.; Goldberg, L. I. *Circ. Res.* 1965, 16, 510.

Joseph G. Cannon,* Basil J. Demopoulos
Division of Medicinal Chemistry and Natural Products
College of Pharmacy

John Paul Long, Jan R. Flynn, Fouad M. Sharabi
Department of Pharmacology, College of Medicine
The University of Iowa, Iowa City, Iowa 52242

Received May 20, 1980